

A Review of the Use of Biotin for Hair Loss

Deepa P. Patel^{a, b} Shane M. Swink^c Leslie Castelo-Soccio^a

^aSection of Pediatric Dermatology, Department of Pediatrics, Children's Hospital of Philadelphia, Philadelphia, PA,

^bUniversity of Louisville School of Medicine, Louisville, KY, and ^cPhiladelphia College of Osteopathic Medicine, Philadelphia, PA, USA

Keywords

Biotin · Hair · Nails · Vitamin · Supplement

Abstract

Background: Biotin has gained commercial popularity for its claimed benefits on healthy hair and nail growth. Despite its reputation, there is limited research to support the utility of biotin in healthy individuals. **Objective:** To systematically review the literature on biotin efficacy in hair and nail growth. **Methods:** We conducted a PubMed search of all case reports and randomized clinical trials (RCTs) using the following terms: (biotin and hair); (biotin and supplementation and hair); (biotin supplementation); (biotin and alopecia); (biotin and nails); (biotin and dermatology), and (biotin recommendations). **Results:** We found 18 reported cases of biotin use for hair and nail changes. In all cases, patients receiving biotin supplementation had an underlying pathology for poor hair or nail growth. All cases showed evidence of clinical improvement after receiving biotin. **Conclusions:** Though its use as a hair and nail growth supplement is prevalent, research demonstrating the efficacy of biotin is limited. In cases of acquired and inherited causes of biotin deficiency as well as pathologies, such as brittle nail syndrome or uncombable hair, biotin supplementation may be of benefit. However, we propose these cases are uncommon and that there is lack of sufficient evidence for supplementation in healthy individuals.

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Introduction

Biotin (also known as vitamin B7 or vitamin H) is a water-soluble vitamin that serves as an essential cofactor for carboxylase enzymes in multiple metabolic pathways. Due to its relatively low cost and abundance of availability in cosmetic products, biotin has become the new trend for consumers wishing to have longer, healthier hair and nails. Current recommendations for biotin by the Institute of Medicine state that the daily adequate intake (AI) for adults is 30 µg/day [1]. Most healthy individuals meet these requirements through a well-balanced diet, though many still take up to 500–1,000 µg of biotin supplementation daily. Although no major toxicities of excess biotin have been reported, data on the actual benefit of biotin's effect on hair and nail growth is limited. Moreover, outside the setting of pregnancy, malnutrition, medication effects, and biotinidase deficiency in children, reports of low biotin levels have rarely been cited. Therefore, we propose that true biotin deficiency is uncommon and that there is lack of sufficient evidence for supplementation for hair and nail growth in individuals who do not present with low levels of biotin.

Table 1. Reported cases of patients categorized by age, dose of biotin, symptoms, and length of treatment until clinical improvement

Study	Age	Reason for biotin deficiency identified	Alopecia reported	Nail changes reported	Dose of biotin	Reported time and degree of hair improvement
Dakshinamurti and Triggs-Raine, 1997 [2]	newborn	yes, inherited enzyme deficiency	no	no	2,500 µg/day	clinical improvement by 6 months of age
Rajendiran and Sampath, 2011 [3]	2 months	yes, inherited enzyme deficiency	yes	no	10,000 µg/twice a day	total resolution in 8 months
Fujimoto et al., 2005 [4]	5 months	yes, dietary/only on amino acid formula for dyspepsia	yes	no	1,000 µg/day	hair regrowth in 2 months
Colamaria et al., 1989 [5]	4 months	yes, inherited enzyme deficiency	not true alopecia, but sparse scalp hair was reported	no	5,000 µg/twice a day	clinical improvement of neurological symptoms seen after 10 days of starting biotin
Coulter et al., 1982 [6]	11 months	yes, inherited enzyme deficiency	yes	no	10,000 µg/day	not specified
Coulter et al., 1982 [6]	14 months	yes, inherited enzyme deficiency	yes	no	10,000 µg/twice a day	not specified
Boccaletti et al., 2007 [7]	1 year	no	no (uncombable hair syndrome)	yes, onychoschizia of nail plates of hands and feet	5,000 µg/day	excellent results in 3 months
Shelley and Shelley, 1985 [8]	18 months	no	no (uncombable hair syndrome)	no	300 µg/3 times daily	significant improvement in 4 months
Boccaletti et al., 2007 [7]	2 years	no	no (uncombable hair syndrome)	yes, onychoschizia of nail plates of hands and feet	5,000 µg/day	excellent results in 3 months
Mukhopadhyay et al., 2014 [9]	3 years	yes, inherited enzyme deficiency	yes	no	30,000 µg/day	6-week follow-up showed dramatic improvement in scalp
Komur et al., 2011 [10]	3 years	yes, inherited enzyme deficiency	yes	no	10,000 µg/twice a day	marked improvement in 1 month, complete hair growth in 6 months
Gannavarapu et al., 2015 [11]	5 years	yes, inherited enzyme deficiency	yes	no	10,000 µg/day	n/a
Rahman et al., 1997 [12]	5 years	yes, inherited enzyme deficiency	yes	no	10,000 µg/day	marked clinical improvement of neurological symptoms was observed at 5 months
Roth et al., 1980 [13]	5 years	yes, inherited enzyme deficiency	yes	no	20,000 µg/day	n/a
Castro-Gago et al., 2011 [14]	n/a	valproic acid, but no significant decreases in biotin and biotinidase levels seen	yes	no	10,000 µg/day	3 months
Hochman et al., 1993 [15]	n/a	yes, brittle nail syndrome	no	yes	2,500 µg/day	6 months
Colombo et al., 1990 [16]	n/a	yes, brittle nail syndrome	no	yes	3,000 µg/day	2 months
Floersheim, 1989 [17]	n/a	yes, brittle nail syndrome	no	yes	2,500 µg/day	n/a

Materials and Methods

We conducted a PubMed search of all case reports and randomized clinical trials (RCTs) published using the following terms: (biotin and hair); (biotin and supplementation and hair); (biotin supplementation); (biotin and alopecia); (biotin and nails); (biotin and dermatology), and (biotin recommendations). We identified additional sources through references contained in the original articles. We limited our search to studies discussing human subjects only. Through this search, we found 18 reported cases of biotin use for hair and nail changes (Table 1).

Of the reported cases in the literature, all patients receiving biotin supplementation had some underlying pathology for either poor hair or nail growth. Moreover, all cases showed evidence of clinical improvement after receiving biotin. Time to improvement as well as dosage administered varied for each case. Ten of the 18 cases were reports of patients with inherited enzyme deficiency in either biotinidase or holocarboxylase synthetase. Of these 10, 8 cases reported alopecia that subsequently resolved after varying months of biotin supplementation. Additionally, there were 3 reported cases of uncombable hair syndrome that all showed improvement in hair quality after a few months of treatment. Fujimoto et al. [4] reported a case of biotin deficiency secondary to diet

Table 2. Established AI levels of biotin per the Food and Nutrition Board of the Institute of Medicine [1]

Life stage	Age	Males, µg/day	Females, µg/day
Infants	0–6 months	5	5
Infants	7–12 months	6	6
Children	1–3 years	8	8
Children	4–8 years	12	12
Children	9–13 years	20	20
Adolescents	14–18 years	25	25
Adults	≥19 years	30	30
Pregnancy	all ages	n/a	30
Breast-feeding	all ages	n/a	35

in an infant who was consuming a special amino acid formula. This patient had low serum and urine levels of biotin as well as perioral dermatitis and alopecia. Hair regrowth in this patient occurred after 2 months of biotin supplementation. Only 1 study conducted by Castro-Gago et al. [14] showed decreased levels of both biotin and biotinidase secondary to medication usage (valproic acid) that improved after 3 months of supplementation with biotin. Finally, 3 cases of brittle nail syndrome treated with biotin were found in the literature and each case showed improvement of nail strength as well as growth on either 2,500 or 3,000 µg of biotin/day.

Discussion

Biotin is a required cofactor for carboxylase enzymes that become activated once they are joined together by holocarboxylase synthase [18]. These enzyme complexes play an important role in multiple metabolic processes including gluconeogenesis, fatty acid synthesis, and amino acid catabolism [19]. Biotin’s function in protein synthesis and more specifically, in keratin production, explains its contribution to healthy nail and hair growth. Biotin is readily found in many foods and is also produced by normal gut flora. Foods found to have high amounts of biotin include nuts, legumes, whole grains, unpolished rice, and egg yolk [20]. Recommended daily allowances of biotin have not been established due to a lack of sufficient evidence [21]. However, AI levels have been recommended by the Food and Nutrition Board of the Institute of Medicine (Table 2). It has been estimated that in Western populations, typical dietary intake of biotin is between 35 and 70 µg/day [22]. Though several animal models [23–25] demonstrating the effects of induced biotin deficiency can be found in the literature, there are currently no studies that show biotin deficiencies in healthy human individuals with balanced diets.

Biotin deficiency can be either acquired or congenital. Though an acquired biotin deficiency is possible, it is still rare. A commonly documented cause of acquired biotin deficiency is secondary to increased raw egg consumption. The protein avidin, found in raw egg whites, can be denatured through cooking, but when uncooked, this protein binds to biotin tightly preventing it from being used as an essential cofactor [26]. Patients taking anticonvulsant medications, such as valproic acid, can also become deficient, and therefore, are prophylactically administered biotin [21]. Additional causes of acquired biotin deficiency include states of alcoholism or pregnancy, other medications, such as isotretinoin [27], impaired intestinal absorption, or prolonged use of antibiotics interrupting normal gut flora [18, 23, 24]. Congenital or genetic biotin deficiency is due to an autosomal recessive trait leading to a lack of either holocarboxylase synthase or biotinidase. When it occurs within the first 6 weeks of life, this deficiency is defined as the neonatal type. In this type of biotin deficiency, the enzyme holocarboxylase synthetase is absent and patients typically have severe, life-threatening conditions [18, 28, 29]. Beyond 3 months of life, the infantile form predominates and is defined by a biotinidase deficiency which is involved in the absorption of free biotin following carboxylase degradation [18, 28, 30]. Whether acquired or congenital, typical symptoms of biotin deficiency include alopecia, eczematous skin rashes, seborrheic dermatitis, conjunctivitis, and multiple neurological symptoms, such as depression, lethargy, hypotonia, and seizures [3, 20]. While the neurological symptoms occur at more severe levels of biotin deficiency, the dermatological manifestations often appear first and are therefore an important indicator [31]. The normal biotin plasma concentration ranges from 400 to 1,200 ng/L [22]. Deficiency is technically considered to be a level of less than 200 ng/L. However, plasma biotin levels can fluctuate daily and thus are not considered to be a sensitive marker [22]. A more validated measure of biotin deficiency is an increased urinary excretion of the metabolite, 3-hydroxyisovaleric acid (normal level: 195 µmol/24 h) [22].

In our search, we found 18 reports in the literature that showed improvement of hair and nail growth on supplementation in patients with established biotin deficiency. For patients with inherited enzyme deficiency, larger doses of biotin supplementation are recommended (from 10,000 to 30,000 µg/day). Those with brittle nail syndrome and other underlying hair pathologies, such as uncombable hair syndrome, require much lower doses of biotin supplementation ranging from 300 to 3,000 µg/

day. Despite these data, there have been no randomized, controlled trials to prove the efficacy of supplementation with biotin in normal, healthy individuals. Moreover, only 1 case in the literature has measured the levels of biotin in normal individuals that had complaints of hair loss. In this study of 541 women (age range between 9 and 92 years), 38% had low biotin levels [20]. However, of those women, 11% were later found through patient history (use of antibiotics, antiepileptics, isotretinoin, or GI disease) to have a reason for the underlying deficiency and 35% had co-existing seborrheic dermatitis, suggesting a multifactorial cause for hair loss. Additionally, in vitro studies have shown that proliferation and differentiation of normal, nonpathologic follicular keratinocytes are not influenced by biotin [27].

Conclusions

Despite its popularity in the media and amongst consumers, biotin has no proven efficacy in hair and nail growth of healthy individuals. Only 1 study has shown decreased levels of biotin in healthy individuals, though this data was confounded by multiple factors, including patient history. Therefore, in the absence of additional studies, we have found no evidence to suggest benefit from biotin supplementation outside of known deficiencies secondary to congenital or acquired causes.

Disclosure Statement

The authors declare no conflicts of interest pertaining to the current publications.

References

- 1 Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes and Its Panel on Folate, Other B Vitamins, and Choline. Dietary Reference Intakes for Thiamin, Riboflavin, Niacin, Vitamin B6, Folate, Vitamin B12, Pantothenic Acid, Biotin, and Choline. Washington, National Academies Press (US), 1998.
- 2 Dakshinamurti K, Triggs-Raine B: Biotin and Multiple Carboxylase Deficiency. Clinical Studies in Medical Biochemistry. Oxford University Press, 1997.
- 3 Rajendiran A, Sampath S: Biotinidase deficiency – clinching the diagnosis rapidly can make all the difference! *BMJ Case Reports* 2011;2011.pii:bcr0720114494. DOI 10.1136/bcr.07.2011.4494.
- 4 Fujimoto W, Inaoki M, Fukui T, et al: Biotin deficiency in an infant fed with amino acid formula. *J Dermatol* 2005;32:256–261.
- 5 Colamaria V, Burlina AB, Gaburro D, Pajno-Ferrara F, Saudubray JM, Merino RG, Bernardina BD: Biotin-responsive infantile encephalopathy: EEG-polygraphic study of a case. *Epilepsia* 1989;30:573–578.
- 6 Coulter DL, Beals TF, Allen RJ: Neurotrichosis: hair-shaft abnormalities associated with neurological diseases. *Dev Med Child Neurol* 1982;24:634–644.
- 7 Boccaletti V, Zendri E, Giordano G, Gnetti L, De Panfilis G: Familial uncombable hair syndrome: ultrastructural hair study and response to biotin. *Pediatric Dermatol* 2007;24:E14–E16.
- 8 Shelley WB, Shelley ED: Uncombable hair syndrome: observations on response to biotin and occurrence in siblings with ectodermal dysplasia. *J Am Acad Dermatol* 1985;13:97–102.
- 9 Mukhopadhyay D, Das MK, Dhar S, Mukhopadhyay M: Multiple carboxylase deficiency (late onset) due to deficiency of biotinidase. *Indian J Dermatol* 2014;59:502–504.
- 10 Komur M, Okuyaz C, Ezgu F, Atici A: A girl with spastic tetraparesis associated with biotinidase deficiency. *Eur J Pediatr Neurol* 2011;15:551–553.
- 11 Gannavarapu S, Prasad C, DiRaimo J, et al: Biotinidase deficiency: spectrum of molecular, enzymatic and clinical information from newborn screening Ontario, Canada (2007–2014). *Mol Genet Metab* 2015;116:146–151.
- 12 Rahman S, Standing S, Dalton RN, Pike MG: Late presentation of biotinidase deficiency with acute visual loss and gait disturbance. *Dev Med Child Neurol* 1997;39:830–831.
- 13 Roth KS, Yang W, Foremann JW, Rothman R, Segal S: Holocarboxylase synthetase deficiency: a biotin-responsive organic acidemia. *J Pediatr* 1980;96:845–849.
- 14 Castro-Gago M, Perez-Gay L, Gomez-Lado C, et al: The influence of valproic acid and carbamazepine treatment on serum biotin and zinc levels and on biotinidase activity. *J Child Neurol* 2011;26:1522–1524.
- 15 Hochman LG, Scher RK, Meyerson MS: Brittle nails: response to daily biotin supplementation. *Cutis* 1993;51:303–305.
- 16 Colombo VE, Gerber F, Bronhofer M, Floersheim GL: Treatment of brittle fingernails and onychoschizia with biotin: scanning electron microscopy. *J Am Acad Dermatol* 1990;23(6 Pt 1):1127–1132.
- 17 Floersheim GL: Treatment of brittle fingernails with biotin. *Z Hautkr* 1989;15:64:41–48.
- 18 Goldberg LJ, Lenzy Y: Nutrition and hair. *Clin Dermatol* 2010;28:412–419.
- 19 Glew RH, Stephen PP: Clinical studies in medical biochemistry. New York, Oxford University Press, 1987.
- 20 Trüeb RM: Serum biotin levels in women complaining of hair loss. *Int J Trichology* 2016;8:73–77.
- 21 Higdon J, Drake VJ: An evidence-based approach to vitamins and minerals. Google books. New York, Thieme, 2012. <http://books.google.com> (accessed March 16, 2010).
- 22 Zemleni J, Mock DM: Biotin biochemistry and human requirements. *J Nutr Biochem* 1999;10:128–138.
- 23 Finner AM: Nutrition and hair deficiencies and supplements. *Dermatol Clin* 2013;31:167–172.
- 24 Zemleni J, Hassan YI, Wijeratne SSK: Biotin and biotinidase deficiency. *Expert Rev Endocrinol Metab* 2008;3:715–724.
- 25 Rushton DH: Nutritional factors and hair loss. *Clin Exp Dermatol* 2002;27:396–404.
- 26 Mock DM: Skin manifestations of biotin deficiency. *Semin Dermatol* 1991;10:296–302.
- 27 Schulpis KH, Georgala S, Papakonstantinou ED, et al: The effect of isotretinoin on biotinidase activity. *Skin Pharmacol Appl Skin Physiol* 1999;12:28–33.
- 28 Miller SJ: Nutritional deficiency and the skin. *J Am Acad Dermatol* 1989;21:1–30.
- 29 Nyhan WL: Inborn errors of biotin metabolism. *Arch Dermatol* 1987;123:1696a–1698a.
- 30 Venkataraman V, Balaji P, Panigrahi D, et al: Biotinidase deficiency in childhood. *Neurol India* 2013;61:411–413.
- 31 Limat A, Suormala T, Hunziker T, Waelti ER, Braathen LR, Baumgartner R: Proliferation and differentiation of cultured human follicular keratinocytes are not influenced by biotin. *Arch Dermatol Res* 1996;288:31–38.

Biotin for the treatment of nail disease: what is the evidence?

Shari R Lipner ¹, Richard K Scher ¹

Affiliations Expand

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Abstract

Aim: To describe the pharmacology, mechanism of action, and clinical reports using biotin to treat nail conditions.

Methods: A review of articles indexed for MEDLINE on PubMed using keywords 'biotin' and 'nail' was performed and applicable articles were selected for review.

Results: Clinical trials have shown an improvement in firmness, hardness, and thickness of brittle nails with oral biotin. There are some case reports and series demonstrating that oral biotin may improve triangular worn down nails, trachyonychia, and habit tic nail deformity.

Conclusions: Oral biotin has been used to treat several nail conditions with promising results. Further larger clinical trials with controls are necessary to determine efficacy and optimal dosing.

Keywords: Nails; biotin; brittle nails; habit tic nail; trachyonychia; triangular worn down nails; vitamins.

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Investigations on *Piper betle* grown in Sri Lanka

Arambewela L. S. R., Arawwawala L. D. A. M., Kumaratunga K. G¹, Dissanayake D. S, Ratnasooriya W. D. ², Kumarasingha S. P.³

Industrial Technology Institute, Baudhaloka Mawatha, Colombo 7, ¹Food Control Laboratory, Ministry of Health, Anuradhapura, ²Department of Zoology, University of Colombo, ³Colombo North Teaching Hospital, Ragama, Sri Lanka (currently at Royal Perth Hospital, Perth, Australia)

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ABSTRACT

Piper betle is an economically important plant cultivated in Sri Lanka. Although more than 12 cultivars of betel are reported in Sri Lanka, very few scientific investigations have been carried out on them. Studies on the chemical constituents indicated that safrole is the major constituent, followed by chavibitol acetate, in the essential oil of common betel leaves of Sri Lanka. Investigations on the bioactivities of *P. betle* revealed the presence of antimicrobial, insecticidal, antioxidant, antinociceptive, antidiabetic and gastroprotective activities. In addition, *P. betle* was found to be safe in terms of hepatotoxicity, renotoxicity, hematotoxicity, gross morphology, weights of organs, stress or aversive behaviors in rats. The above findings indicate the vast potential of *P. betle* yet to be harnessed for the benefit of mankind and the betel industry of Sri Lanka.

Key words: Anatomy, bioactivities, chemical constituents, morphology, *Piper betle*, safety profile

INTRODUCTION

Piper betle Linn. (Sinhala name: Bulath, English name: Betel) belongs to the genus *Piper* of the plant family Piperaceae.^[1] Over 700 species of plants belonging to the genus *Piper* are distributed in both hemispheres.^[2] Of these, about 30 species have been reported from India.^[1] In Sri Lanka, 18 species of genus *Piper* are found; and out of them, three are endemic.^[3]

Betel leaves have been traditionally used for chewing purposes along with other condiments. Colombo, Gampaha, Kalutara, Kurunegala, Kegalle, Ratnapura, Matale and Galle are the main betel-cultivating districts in the country. In addition to a wide and well spread domestic market, betel has gained a significant position in the export market since 1974. Betel industry, at times, faces severe problems of depressed prices and restricted export market. The main cause for this situation is that Pakistan, our

major buyer of betel, has reduced the volume of betel imported from Sri Lanka.^[4]

Although betel vine has been cultivated in Sri Lanka for centuries, very few research activities have been carried out on it, except studies on antiaphrodisiac activity,^[5] antifertility effects on male rats^[6] and antimotility effects on washed human spermatozoa.^[7] However, *P. betle* grown in other countries has been shown to possess antimicrobial,^[8] gastroprotective,^[9] wound healing,^[10] hepatoprotective^[11] and antioxidant^[12] activities. In order to minimize the negative impact on the betel industry in Sri Lanka, there is a necessity to study important bioactivities of Sri Lanka grown betel leaves and develop value added products based on these activities. This strategy will safeguard both growers and the economy of the country. In order to achieve this, possible bioactivities of betel oil and extracts were investigated using betel leaves from Sri Lanka. In addition, morphological, anatomical and chemical studies were also conducted. These investigations and their results are summarized below.

Address for correspondence:

Dr. Menuka Arawwawala
Industrial Technology Institute, Baudhaloka Mawatha, Colombo 7, Sri Lanka. E-mail: menukaarawwawala@yahoo.com/
menuka@iti.lk

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CLASSIFICATION

Kingdom: Plantae
Order: Piperales
Family: Piperaceae
Genus: Piper
Species: *P. betle*

Morphological and anatomical studies of betel

Six cultivars of *P. betle* Linn., namely, Galdalu, Mahamaneru, Kudamaneru, Ratadalu, Nagawalli and Malabulath, were used

in the present study.^[13] Morphological and anatomical features, including parameters such as stomatal index, leaf length-to-width ratio, were similar in Kudamaneru, Mahamaneru, Galdalu, Ratadalu and Nagawalli but were different in the cultivar Malabulath.

CHEMICAL CONSTITUENTS AND PHYSICOCHEMICAL PROPERTIES OF THE ESSENTIAL OIL

Chemical composition

According to the chemical constituents present in the essential oil (EO), cultivars of Galdalu, Mahamaneru, Kudamaneru, Ratadalu and Nagawalli were similar and they were categorized under “common betel.” EOs from the leaves, stalks, stems, fruits and roots of common betel and from the leaves of Malabulath were also analyzed.^[13] Major constituents of the EO of common betel were found to be safrole (48.7%) and chavibitol acetate (12.5%). Malabulath does not contain these two compounds. The major compound in Malabulath oil is allylpyrocatechol diacetate (34.0%), which is the third major compound in common betel oil (11.3%). Further, *p*-cymene, 4-terpineol, safrole, eugenol, β -caryophellene and chavibitol acetate detected in common betel leaf oil were not detected in Malabulath leaf oil. The Gas Chromatography-Mass Spectroscopy analysis of the EO of different parts of common betel indicated that composition of the stalk oil was different to that of the other parts, as it did not contain detectable amounts of allylpyrocatechol diacetate. The major compound detected in the EO from the leaf, the stem, the stalk and the root was safrole; but in the fruit oil, it was β -phellandrene. This chemical composition of the EO of leaves appears to be closer to that of cultivar *Deshawari* in India.

The composition of the betel EO changes with the maturity of the leaf. It was observed that the contents of major compounds safrole and chavibitol acetate in the leaf were at the maximum level at the harvesting stage. Moreover, eugenol and β -phellandrene content decreased with maturity, and β -phellandrene content remained constant after maturity. Allylpyrocatechol diacetate content increased up to the harvesting stage and remained constant thereafter. The study on the variation of the composition of the EO with maturity is useful in deciding the maturity stage at which the leaf has to be collected for applications that depend on specific compounds. Further, it justifies why Ayurvedic physicians mention the maturity of the plant in drug preparations.

Physicochemical properties

The physicochemical properties of the EOs of Kudamaneru, Mahamaneru, Galdalu, Ratadalu and Nagawalli too were similar but were different from those of Malabulath. These studies indicate that physicochemical properties and chemical constituents of the EO of Malabulath are different from those of other cultivars.

ANTIMICROBIAL SCREENING STUDIES

Antibacterial activity

In the present study, the EO from the leaves showed activity against *Escherichia coli*, *Streptococcus pyogenes* and *Staphylococcus aureus*. The Minimum Inhibitory Concentration (MIC) values were 3.12×10^2 , 2.50×10^3 and 5.00×10^3 $\mu\text{g/mL}$, respectively. The ethanol extract showed activity against *Streptococcus pyogenes*, *Escherichia coli* and *Staphylococcus aureus*. The MIC values were 1.25×10^3 , 5.00×10^3 and 5.00×10^3 $\mu\text{g/mL}$, respectively.^[13, 14]

Antifungal study

This assay revealed that EO of *P. betle* contained at least three fungicidal compounds, and the ethanol extract contained at least one fungicidal compound active against *Cladosporium sp.* Further, antifungal activity of betel oil was investigated against *Colletotrichum sp.*, *Fusarium oxysporum sp.*, *Corynospora cassicola*, *Rigidoporus sp.* and *Phytophthora sp.*, using the disk method. All fungi species except *Phytophthora sp.* showed significant growth inhibition in betel oil.^[13, 14]

INSECTICIDAL ACTIVITIES

Mosquito larvicidal assay

Late 3rd instar larvae of *Aedes egyptii* were introduced into *P. betle* EO at 500, 100, 50, 25, 12.5, 6.25 ppm concentrations, and mortalities were recorded after 1 hour and 24 hours. Mortalities of 43% and 100% were observed for 100 and 500 ppm concentrations, respectively, within 1 hour. Compared to the control, significant mortality was observed even at lower concentrations, 25 and 50 ppm, after 24 hours.^[14]

Bioassay for housefly (*Musca domestica*)

In this assay, betel EO (120 $\mu\text{g/cm}^3$ in ethanol) showed 100% knock down effect and mortality against *Musca domestica*.^[14]

Bioassay for rice weevil (*Sitophilus oryzae*)

Betel EO at 1%, 0.8% and 0.5% concentrations was used in this study. Mortality rate of 100% was observed in 1% betel oil solution within 1½ hours.^[14]

Bioassay for *Chrysomya megacephala* larvae

Betel EO solutions ranging in concentration from 1% to 4% were prepared using 1% Tween 80, sodium lauryl sulfate (0.05 g per 100 mL, as a stabilizer) and methyl paraben (0.01 g per 100 mL, as a preservative). The 4% and 3% preparations of the oil of betel were effective in killing 100% of the larvae of *C. megacephala* within 3½ hours, while betel oil at 2% concentration killed 97% of *C. megacephala* larvae within 4 hours. The positive control, mineral turpentine, also killed the larvae within 4 hours. This shows that betel oil is effective in the treatment of wound myiasis.^[15]

Bioassay for *Chrysomya bezziana* larvae

A study was conducted to evaluate the efficacy of betel EO against the larvae of *Chrysomya bezziana* *in vitro*.^[10] With 4% betel

oil, all 1st instar larvae were killed within 2 hours, and the 2nd instar larvae were killed within 4 hours. The positive control (Asuntol) showed no mortality until 4 hours, but all larvae were weak - from first 30 minutes. In the negative control, larvae were mobile and active. Betel oil at 3% killed all the 1st instar larvae within 150 minutes; and 74% of the 2nd instar larvae, within 4 hours. These results indicate that betel oil from Sri Lanka is an effective larvicide for *C. bezgiana* 1st and 2nd instar larvae *in vitro*.

ANTIOXIDANT ACTIVITY

The extracts obtained from the leaves of *P. betle* had profound antioxidant activity as judged by Thiobarbituric Acid Reactive Substances (TBARS) and 2, 2 – diphenyl – 1 – picrylhydrazyl (DPPH) scavenging assays.^[14,17] The scavenging effects of *P. betle* extracts on DPPH radicals increased in the following order: Cold Ethanolic Extract (CEE) > EO > HWE. Further, free radical scavenging effect of CEE was higher than that of synthetic antioxidant Butylated Hydroxy Toluene (BHT). Employing the DPPH assay, Indian researchers^[12] have investigated the antioxidant activities of three betel cultivars (Kauri, Ghanagete and Bagerhati) grown in India. The antioxidant activities of the three cultivars were in the order of Kauri > Ghanagete > Bagerhati, but the free radical scavenging ability of commercial betel from Sri Lanka is higher than that of the reported Indian cultivars.

In TBARS assay, the antioxidant potential of CEE was the best among the *P. betle* extracts tested, and this effect was significantly higher than that of BHT and green tea, respectively. Compared to EO and HWE, the degree of delaying the lipid peroxidation was significantly lower in BHT and in green tea. Safrole is the major constituent in the EO of Sri Lankan commercial betel leaves. However, compared to the EO, the antioxidant activity of safrole was significantly low. This suggests that antioxidant potential of EO is not only due to safrole but possibly due to synergetic effect of all volatile constituents.

Interestingly, the antioxidant properties of *P. betle* extracts, including CEE, EO and HWE, remained unaltered for a period of 12 months at room temperature (as evaluated by DPPH assay). This supports the potential use of the betel extracts as a natural antioxidant in food industry. However, when *P. betle* extracts were exposed to elevated temperature (200°C), the antioxidant property was significantly reduced (CEE and EO by 4 fold; HWE by 3 fold). Similar results were also evident with the synthetic antioxidant BHT (by 4 fold). Even after exposure to the elevated temperature, the antioxidant potential of CEE was higher than that of BHT.

In an attempt to introduce betel as a natural antioxidant, the CEE was incorporated into fats (cake margarine), oils (coconut and palm oil) separately and its rancidity determined in terms of peroxide value (PV). The results showed that PVs were significantly lower in CEE-treated samples than in BHT-treated samples.

ANTIDIABETIC ACTIVITY

Overall results show that both HWE and CEE of *P. betle* leaves from Sri Lanka possess marked hypoglycemic activity (when tested in fasted normoglycemic rats) and antihyperglycemic activity (judged by improvement in the results of glucose tolerance test and by lowering of the blood glucose levels in rats with streptozotocin (STZ)-induced diabetes).^[18] The hypoglycemic effect of *P. betle* extracts (100, 200, 300 mg/kg) on fasted normoglycemic rats was dose dependent and lasted up to 4 hours, except that of the lowest dose of HWE. Further, hypoglycemic potential of HWE and CEE, respectively, was comparable to that of tolbutamide, the reference hypoglycemic drug of the sulphonylurea type.

In glucose tolerance test, HWE, CEE and tolbutamide lowered the external glucose level in a similar manner. Further, HWE significantly reduced the blood glucose level of rats with STZ-induced diabetes treated with a dose (50 mg/kg) which is known to irreversibly damage the insulin-secreting β cells of the pancreas. This suggests that an intact-endocrine pancreas and insulin are not essential for antidiabetic activity of *P. betle* extracts. This ability of lowering the blood glucose levels of rats with STZ-induced diabetes also suggests that *P. betle* extracts have insulinomimetic activity. It is possible that HWE may act as an insulin secretagogue and/or sensitize insulin receptors, as proposed for some plant extracts. HWE failed to significantly inhibit glucose absorption from the lumen of the intestine. However, HWE provoked accumulation of glycogen in the liver and the skeletal muscle. This is another peripheral mechanism through which HWE exhibits its antidiabetic activity. This increased glycogenesis may result from enhanced glucose uptake from liver and skeletal muscle by sensitization of insulin receptors and/or inducing the activity of enzymes involved in glycogen synthesis.

GASTROPROTECTIVE ACTIVITY

A study to evaluate the gastroprotective activity of HWE and CEE of *P. betle* leaves was carried out.^[19] Three doses (200, 300 and 500 mg/kg) of both the extracts were evaluated for gastroprotective activity against ethanol-induced gastric ulcers in rats. Oral administration of HWE and CEE provided marked dose-dependent and significant protection against gastric damage caused by absolute ethanol. The gastroprotective effect of HWE was comparable to that of CEE. Further, the gastroprotective activities of the highest dose of both extracts were significantly greater than the gastroprotective activity of misoprostol, the reference drug. A similar study conducted using Indian betel cultivars has been reported.^[9] In this experiment, treatment with ethanol extract of betel leaves at a dose of 150 mg/kg for 10 consecutive days produced significant healing effect in rats with ulcers induced by nonsteroidal anti-inflammatory drugs (NSAIDs).

The HWE significantly increased the mucus content (by 49%)

adhering to the wall of the gastric mucosa. Mucus layer is considered to be important in the mucosal defense against endogenous aggressors, e.g., acids, and also as an agent in facilitating its repair. It is generally believed that enhanced acid secretion is the most important factor for the induction of gastric lesions. In this study, the highest dose of HWE did not cause significant inhibition in acidity (both total and free) or pH of gastric fluid. Therefore, the gastroprotective effect of *P. betle* was not mediated *via* inhibition of acid secretion in the gastric mucosa but by increasing its mucus content.

ANTINOCICEPTIVE ACTIVITY

The CEE and HWE of betel leaves have antinociceptive activity, as evaluated in the hot plate test and in the tail flick test in rats. This indicates centrally mediated antinociceptive activity of the plant extracts against acute pain.^[20] The antinociceptive activity of extracts (100, 200, 300 mg/kg) was genuine, as there was no change in the retain retention times in the Bar and Bridge tests as compared to controls. Both 200 and 300 mg/kg doses of *P. betle* extracts markedly reduced the licking time in early and late phases of the formalin test in a bell-shaped dose-response curve. In the formalin test, the pain in the early phase is caused due to the direct stimulation of the sensory nerve fibers by formalin, while the pain in the late phase is due to the inflammatory mediators, like histamine, prostaglandin, serotonin and bradykinin. It is reported that NSAIDs reduce both phases of the formalin test. The betel extracts too induced interruptions in both phases of this test, suggesting possible impairments of sensory transmission and release of inflammatory mediators. The highest antinociceptive activity was evident with 200 mg/kg dose of both HWE and CEE. As antinociceptive activity of CEE was higher than that of HWE, CEE was used to investigate its antinociceptive mechanism.

Some sedatives possess antinociceptive activity. However, the antinociceptive action of CEE is unlikely to be mediated *via* sedation, as none of the parameters monitored in the Rat hole board technique were changed. Antinociception can be induced via dopaminergic mechanisms, but such a mode of action is unlikely in this study as metoclopramide, a dopamine receptor antagonist, failed to block antinociception induced by the extract. The opioid receptor, antagonist naloxone, blocked the antinociception induced by CEE, suggesting that the antinociception was mediated through opioid mechanisms.

SAFETY PROFILE

There were no treatment-related deaths or morbidity with CEE (1,500 mg/kg/d) and HWE (1,500 mg/kg/d) even after following sub-chronic (14 days) and chronic (42 days) oral treatment in rats.^[21] Further, CEE and HWE treated rats showed normal food intake, water intake, and their percentage weight gain significantly reduced. The consistencies of feces and color of urine in CEE and HWE treated rats were essentially similar to those of the

controls. Furthermore, neither extract of *P. betle* induced any overt signs of toxicity (salivation, diarrhea, lacrimation, tremors, ataxia, yellowing of hair, loss of hair, postural abnormalities or behavioral changes), stress (fur erection or exophthalmia) or aversive behaviors (biting paw and penis, intense grooming behavior, scratching behavior, licking at tail or vocalization).

Neither extract significantly changed any of the serum parameters [aspartate aminotransferase (AST), alanine aminotransferase (ALT), urea and creatinine] and hematological [red blood cell (RBC) counts, white blood cell (WBC) counts and hemoglobin (Hb) concentration] parameters investigated. All the tested organs (liver, kidneys, testes, adrenal glands, heart, spleen, vasa deferentia, prostate glands, seminal vesicles together with coagulating glands and caput plus corpus epididymides) appeared normal in all treated rats. There were no significant changes in the organ weights between the treated groups except for the spleen. Compared to the control, in both treated groups significant increase in weight of the spleen was evident (CEE- 217.4%; HWE- 234.8%). Gastric lesions were not observed in any of the treated rats. In conclusion, this study shows that both cold ethanolic and hot water extracts of Sri Lankan betel leaves and leaf stalk were safe following sub-chronic oral administration to rats.

A similar study conducted in India also reported that ethanolic extract of *P. betle* leaf stalks was nontoxic as judged by hematological, biochemical profiles and enzymatic studies.^[22]

DEVELOPMENT OF VALUE-ADDED PRODUCTS FROM BETEL

Based on the results of scientific investigations, several value-added products such as betel toothpaste, mouthwash, face cream, shampoos, instant betel quid, betel pellet, antitick lotion, antitick powder and wound healing creams were developed in order to enhance the marketability of betel and improve the prospects of the industry. Clinical trial conducted using the wound healing cream on dermatitis patients revealed that treatment was significantly effective on skin rashes.^[23] At present a clinical study is in progress to evaluate the antidiabetic activity of spray-dried powder of betel hot water extract.

CONCLUSIONS

P. betle is a common plant in Sri Lanka, and it can be easily cultivated in any part of the country. This scientific study revealed for the first time the chemical constituents and multifaceted activities of betel cultivated in Sri Lanka. The *Chrysomya megacephala* and *C. bezziana* larvicidal activities of betel as well as its antidiabetic and antinociceptive activities have been reported for the first time, herein. The fact that betel leaves have multiple activities such as antimicrobial, insecticidal, antioxidant, antinociceptive, gastroprotective and antidiabetic, as revealed in this study, indicates that *P. betle* is a good candidate

for future herbal drug preparations and development. This hitherto untapped vast potential of betel grown in Sri Lanka, if properly harnessed, will safeguard the betel industry of Sri Lanka, enhance the livelihood of a large number of villagers depending on betel industry and introduce novel herbal products and drug preparations into the market.

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REFERENCES

1. Traditional Asian Medicines and Natural Products. *Piper* Linn. (Piperaceae) [monograph on CD – ROME] diskD2. Wealth Asia. Asian Health Environmental and Allied Database. 1997.
2. Parmar, VS, Taneja, P, Jha, A, Tyagi, OD, Prasad, AK, Wengel, J, et al. Phytochemistry of genus *Piper*. *Phytochemistry* 1997;46:597-673.
3. Dassanayake MD, Fosberg FR. A Revised Hand Book to the flora of Ceylon. Washington: Publication of Smithsonian Institute and National Science Foundation; 1987.
4. Anonymous. Betel industry of Sri Lanka, present problems and future prospects. Sri Lanka: Publication of Economic research unit, Department of Export Agriculture; 1987.
5. Ratnasooriya WD, Premakumara GA. *Piper betle* leaves impairs masculine sexual behavior of rats. *Med Sci Res* 1996;24:303-6.
6. Ratnasooriya WD, Premakumara GA. *Piper betle* leaves reversibly inhibits fertility of male rats. *Vidyodaya J Sci* 1997;7:15-21.
7. Ratnasooriya WD, Jayawardena KG, Premakumara GA. Antimotility effects of *Piper betle* (L) leaf extract on washed human spermatozoa. *J Natn Sci Coun Sri Lanka* 1990;18:53-60.
8. Nair R, Chanda S. Antimicrobial activity of *Terminalia catappa*, *Manilkara zapota* and *Piper betle* leaf extract. *Indian J Pharmaceut Sci* 2008;70:390-3.
9. Majumdar B, Chaudhuri SG, Ray A, Bandyopadhyay SK. Effect of ethanol extract of *Piper betle* Linn leaf on healing of NSAID - induced experimental ulcer - A novel role of free radical scavenging action. *Indian J Exp Biol* 2003;41:311-5.
10. Santhanam G, Nagarajan S. Wound healing activity of *Curcuma aromatica* and *Piper betle*. *Fitoterapia* 1990;61:458-9.
11. Saravanan R, Prakasam A, Ramesh B, Pugalendi KV. Influence of *Piper betle* on hepatic marker enzymes and tissue antioxidant status in ethanol - treated wister rats. *J Med Food* 2002;5:197-204.
12. Dasgupta N, De B. Antioxidant activity of *Piper betle* L. leaf extract *in vitro*. *Food Chem* 2004;88:219-24.
13. Arambewela L, Kumaratunga KG, Dias K. Studies on *Piper betle* of Sri Lanka. *J Natn Sci Foundation Sri Lanka* 2005;33:133-9.
14. Chemical studies on *Piper betle* and development of value added products. Final report submitted to CARP. [CARP library] (2004).
15. Kumarasingha, SP, Ihalamulla RL, Arambewela LSR, Dissanayake DS. Larvicidal effects of mineral turpentine, low aromatic white spirits, aqueous extracts of *Cassia alata* and aqueous extracts, ethanolic extracts and essential oil of betel leaf (*Piper betle*) on *Chrysomya megacephala*. *Int J Dermatol* 2002;41:877-80.
16. Wardhana AH, Kumarasingha SP, Arawwawala LDAM, Arambewela LSR. Larvicidal efficacy of essential oil of betel leaf (*Piper betle*) on the larvae of the old world screwworm fly, *Chrysomya bezziana* *in vitro*. *Indian J Dermatol* 2007;52:43-7.
17. Arambewela L, Arawwawala M, Rajapaksa D. *Piper betle*: A natural antioxidant. *Int J Food Sci Technol* 2006;41:10-4.
18. Arambewela LSR, Arawwawala LDAM, Ratnasooriya WD. Antidiabetic activities of aqueous and ethanolic extracts of *Piper betle* leaves in rats. *J Ethnopharmacol* 2005;102:239-45.
19. Arambewela LSR, Arawwawala LDAM, Ratnasooriya WD. Gastroprotective activities of Sri Lankan *Piper betle* leaf extracts in rats. SLAAS. 60th Annual Session. 2004. p. 117.
20. Arambewela LSR, Arawwawala LDAM, Ratnasooriya WD. Antinociceptive activities of aqueous and ethanolic extracts of *Piper betle* leaves in rats. *Pharmaceut Biol* 2006;43:766-72.
21. Arambewela LSR, Arawwawala LDAM, Ratnasooriya WD. Safety evaluation of Sri Lankan *Piper betle* leaf extracts in rats. *J Trop Med Plants* 2003;4:195-8.
22. Sengupta A, Adhikary P, Bask BK, Chakrabarti K, Gangopadhyay P, Banerji J, et al. Pre - clinical toxicity evaluation of leaf stalk extractive of *Piper betle* Linn. in rodents. *Indian J Exp Biol* 2000;38:338-42.
23. Arambewela LSR, Arawwawala LDAM, Withanage D, Kulathunga S. Efficacy of betel cream on skin ailments. *J Complement Integr Med* 2010; 7, Article 48.

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